SL2 cells (Drosophila melanogaster)

SL2 cells were derived from embryonic stem cells and are routinely cultured in Schneider's medium supplemented with 10% FCS, glutamine and antibiotics at 26°C and at density of 3-8 x 10^6 cells/ml. Cells were seeded at day of transfection into 6-well plates at a density of about 5 x 10^5 cells per 2 ml complete medium and were left undisturbed for 1h adhere. Alternatively, cells were seeded 1 day prior to transfection to reach 50-80% confluency after 18-24 hours. For each transfection, 2 µg of DNA and 12 µg of INSECTOGENE were diluted each into 100 µl serum-free medium, mixed gently and incubated at room temperature for 15 min in a 15 ml polystyrene tube. Cells were washed once with serum-free medium and overlaid with the DNA/INSECTOGENE mixture prior diluted into 0.8 ml serum-free medium. Following incubation of 4-6 hours, 1 ml of medium containing 10 % FCS was added. Cells were harvested 18 hours posttransfection and assayed for Luciferase and ß-galactosidase activity luminometically. Luciferase and ß-galactosidase activities were determined as the mean of four independent experiments. Each sample was measured in duplicates.

Obtained efficiency was between 30-35 % (RLU ca. 1.2-1.4 x 10^7)